International Journal of Radiology and Diagnostic Imaging



E-ISSN: 2664-4444 P-ISSN: 2664-4436 www.radiologypaper.com IJRDI 2023; 6(4): 08-13 Received: 13-06-2023 Accepted: 22-07-2023

Farah Kasim Najee Babylon Health Directorate, Babylon, Iraq

Haider N AL-Tameemi

Faculty of Medicine, University of Kufa, Al-Najaf, Iraq

Evaluation of placental stiffness in normal versus high risk pregnancy using shear wave elastography

Farah Kasim Najee and Haider N AL-Tameemi

DOI: https://doi.org/10.33545/26644436.2023.v6.i4a.355

Abstract

Background: Ultrasound is essential for assessing normal and high-risk pregnancies. A unique ultrasonographic method, shear wave elastography (SWE), measures soft tissue component elasticity. This approach has been used in obstetrics lately. This research uses SWE to assess placental stiffness in healthy and high-risk pregnant women.

Method: The Ultrasound clinic of Al-Zahraa teaching hospital in Al-Najaf governorate performed a case-control research of 100 singleton pregnant women (40 cases and 60 controls) from October to December 2022. Cases were pregnant referred from obstetric clinics with gestational hypertension or diabetes, whereas 60 controls were healthy pregnant with no clinical or sonographic signs of high risk. All cases and controls were 2^{nd} or 3^{rd} trimesters. All patients had B-mode ultrasonography and placental SWE exams, and SPSS was used to analyze the data.

Results: There was a significant difference in mean placental SWE values between studied groups, with the highest means found in pregnant with gestational hypertension (2.05 m/s) and gestational diabetes (1.5 m/s) and the lowest mean found in normal pregnant (1.1 m/s), with a cut-off value of 1.27 m/s to distinguish normal and abnormal placenta In high-risk pregnant women, placental thickness, amniotic fluid index, and stiffness were positively correlated.

Conclusion: Normal pregnancy had less placental stiffness than high-risk pregnancy (hypertension and diabetes). Thus, SWE technology may quantify placenta morphological disorders in hazardous pregnant women.

Keywords: Placenta, gestational diabetes, gestational hypertension, shear wave elastography

Introduction

The placenta is vital for fetal and maternal health and affects long-term wellbeing. Research into its development is challenged by ethical concerns, limited in vitro models, and species diversity ^[1, 2]. Ultrasound, a non-ionizing modality, is standard for imaging the placenta, identifying its echogenicity, and distinguishing normal features from pathologies like hematoma^[3]. Recent imaging advancements include elastography, which measures tissue elasticity and is used in assessing organ health with both ultrasound (US) and magnetic resonance (MR) imaging ^[4]. Ultrasound elastography techniques are categorized by the physical quantity measured: strain imaging, including Strain elastography (SE) and Acoustic Radiation Force Impulse (ARFI) imaging, and Shear Wave Imaging (SWI), which includes transient, point, and two-dimensional methods ^[4]. In vivo and ex vivo studies show variable placental elasticity due to different technologies used. High reliability of in vivo measurements has been reported, with consistent Shear Wave Velocity (SWV) values across studies using similar systems ^[5-7]. Factors affecting measurement quality include sample depth and transducer pressure, which may be mitigated by standardized protocols [8-10]. There's limited consensus on normal elasticity at specific gestational stages, and current research shows little significant change in SWV with advancing gestational age [11, 12]. Regional elasticity within the placenta appears consistent across various studies, although some report variability [13, 14]. Confounders like maternal age, blood pressure, and BMI need further investigation [15]. One study suggests increased elasticity with higher BMI, but its reliability is questionable due to large SWV variations ^[16]. Elastography has been used to study conditions like gestational diabetes, showing higher mean shear values compared to normal controls ^[14]. Safety concerns for elastography in pregnancy focus on potential tissue displacement effects from radiation force pulses ^[17, 18]. No placental histological changes have been reported after ARFI imaging ^[5].

Corresponding Author: Farah Kasim Najee Babylon Health Directorate, Babylon, Iraq While elastography uses higher thermal indices, they are within safety limits set by the AIUM ^[19]. Professional bodies like the BMUS and WFUMB endorse the safety of these methods but call for more research to uphold the ALARA principle ^[20]. The aim of study is to measure the placental stiffness in normal and risky pregnancy using the SWE technique and to assess the factors affecting placental stiffness.

Method

A case control study. The data collection was conducted between the first of October to the end of December of 2022 among pregnant women attending the ultrasound clinic in Al-Zahraa teaching hospital in Al- Najaf governorate. We used a sequential sample throughout data gathering. This research recruited second- and third-trimester singleton pregnant women referred by Obstetric clinics to ultrasonography clinics. This research comprised 60 singleton pregnant women with clinically normal and 40 with clinically hazardous pregnancies. This study aimed to evaluate the use of elastography, a technique that measures the stiffness of tissue, in the context of prenatal care. Participants were selected based on stringent criteria: Those with normal physical examinations, laboratory tests, and ultrasound (US) results were categorized as normal pregnant women, while those with histories of gestational diabetes mellitus (DM) and gestational hypertension were considered high-risk. Exclusion criteria were extensive, filtering out pregnancies complicated by fetal congenital anomalies, significant maternal pathologies, placentas with challenging locations or abnormalities, severe maternal anemia, heart disease, or other significant diseases. The research gathered participant data via a two-part questionnaire that included elastography readings, US results, age, and obstetric history. Using a convex transducer, an advanced GE LOGIC E9

XDClear system was used to do ultrasound examinations. Shear wave elastography (SWE) was used to quantify placental stiffness in these assessments in addition to traditional B-mode imaging, which was used to record foetal and placental parameters such as biparietal diameter, femur length, amniotic fluid index, placental location, thickness, and structure. Certain precautions made sure that the patient's respiration and movement did not interfere too much with SWE. In order to determine the velocity values coded S1 through S4 and P, respectively, the placenta was separated into areas and many measurements were made at various locations, including the maternal surface, central portion, foetal surface, and peripheral placenta. Using ANOVA and T-tests for continuous variables, correlation tests for continuous variable connections, chi-square tests for categorical variable associations, and descriptive statistics were used to analyze the data from these measures using SPSS software. With coefficients ranging from mild (0.2-0.29) to extremely high (≥ 0.7) , the correlation strength was assessed. The Receiver Operating Characteristic (ROC) curve was used to evaluate the elastography test's accuracy; areas under the curve classified the test's diagnostic accuracy from excellent to fail. For statistical significance, a P-value of 0.05 or less was used. The rigorous methodology and detailed data collection aimed to establish the reliability and diagnostic utility of elastography in monitoring placental health and potential risks in pregnancy.

Results

A total of 100 pregnant women were enrolled in this study, 60 women with no risk factors and 40 women with risk factor (20 women with gestational diabetic mellitus and 20 women with gestational hypertension). The age distribution between two group were shown in table 1 and the two groups was homogenous regard age (p>0.05).

Table 1:	Age	Distribution	between	two	studied	groups
Table I.	1150	Distribution	bet ween	1110	studied	Sloups

Variables		Partic	P value	
		Normal pregnancy Risky pregnancy		
	< 20 years	6 (10%)	0	
1 22	20-30 years	31 (51.7%)	18 (45%)	
Age	>30 years	23 (38.3%)	22 (55%)	0.058*
	Mean ±SD	27±6.2	31±5.8	
Total		60 (100%)	40 (100%)	

*Chi-Square test, significant ≤0.05.

The obstetric history of studied participants shown that 40% (24) of normal pregnancy women and 62.5% (25) of risky pregnant women had more than four gravidities and there was a significant difference in gravidity and parity between

two studied group ($p \le 0.05$) and there was no significant difference in abortion history between two group (p=0.79), table 2.

Fable 2: Obstetrical history	between	studied	groups.
------------------------------	---------	---------	---------

Variables		Participants		D I
Va	ariables	Normal pregnancy	Risky pregnancy	P value
	Pimi	7 (11.7%)	0	
Gravidity	2-4	29 (48.3%)	15 (37.5%)	0.02*
	>4	24 (40%)	25 (62.5%)	
	Nil parity	Darity 10 (16.7%)	0	
Parity	1-3	34 (56.7%)	21 (52.5%)	0.008*
	>3	16 (26.7%)	19 (47.5%)	
	0	34 (56.7%)	20 (50%)	
Abortion	1-3	25 (41.7%)	19 (47.5%)	0.79*
	>3	1 (1.7%)	1 (2.5%)	
	Total	60 (100%)	40 (100%)	

*Chi-Square test, significant ≤0.05.

There was no significant difference in the placental thickness, gestational age and amniotic fluid index between two studied group (p>0.05). Polyhydromenous was found in risky pregnancy only (15%) and 35% of risky pregnancy

and 21.7% of normal pregnancy had oligohydromenous with a significant difference in the amount of liquor between two studied groups (p=0.001), table 3.

Fable 3: Ultrasound	finding amo	ong two g	roups.
----------------------------	-------------	-----------	--------

Ultrasound finding		Participants		Dreha
		Normal pregnancy	Risky pregnancy	P value
Placenta	l thickness (Mean± SD)	3.6 ±0.75	3.7 ± 0.72	0.669*
Gestational age	BPD (Mean± SD)	231±38	240 ±24	0.21*
	FL (Mean± SD)	238± 36	242 ±25	0.52*
Amount of liquor	Normal	47 (78.3%)	20 (50%)	
	Oligohydromenous	13 (21.7%)	14(35%)	0.001**
	Polyhydromenous	0	6(15%)	
A	AFI (Mean± SD)	15.3 ±4.1	16 ±6.7	0.56*

*Student T test, **Chi-Square test, significant ≤0.05.

There was a significant difference in the mean SWE between studied groups, were the highest mean was found among pregnant with gestational hypertension and lowest

mean was found among normal pregnant (p=0.003), figure 1.



Fig 1: Difference in the mean SWE between two studied groups

Among normal pregnant women. There was no significant correlation between SWE and age of pregnant women and

obstetric history (p>0.05), table 4.

Table 4: Correlation between age and obstetric history with SWE in normal pregnant women.

Variables	SWE Correlation coefficient	P value
Age	0.21	0.87*
Gravidity	0.1	0.41*
Parity	0.5	0.67*
Abortion	0.17	0.19*

Significant ≤0.05.

There was no significant correlation between SWE and ultrasound finding regard placental thickness, BPD, FL and

AFI (p>0.05), table 5.

 Table 5: Correlation between ultrasound finding and SWE among normal pregnant women

Veriables	SWE	Develope
variables	Correlation coefficient	P value
Placental thickness	-0.18	0.155*
BPD	-0.038	0.78*
FL	0.052	0.71*
AFI	-0.11	0.37*

Significant ≤0.05.

https://www.radiologypaper.com

Among the risky women. There was no significant correlation between age, gravidity and parity with SWE, but there was moderate positive correlation between number of abortion and SWE (r=0.32, p=0.039), table 6.

 Table 6: Correlation between age and obstetric history with SWE in risky pregnant women.

Variables	SWE	Dualua
variables	Correlation coefficient	- r value
Age	-0.34	0.053*
Gravidity	0.22	0.169*
Parity	-0.6	0.7*
Abortion	0.32	0.039*
Significant <0.05		

Significant ≤ 0.05 .

There was a significant strong positive correlation between placental thickness, Amniotic fluid index and SWE (r=0.49, 0.59. p=0.001, <0.001 respectively), while there was no significant correlation between BPD and FL and SWE

Table 7: Correlation	between	Ultrasound	finding and	SWE in

risky	pregnant	women.
-------	----------	--------

SWE	Dualma	
Correlation coefficient	r value	
0.49	0.001*	
-0.25	0.166*	
-0.27	0.14*	
0.59	< 0.001*	
	SWE Correlation coefficient 0.49 -0.25 -0.27 0.59	

Significant ≤0.05.

(p>0.05), table 7.

ROC cure and analysis shown that the area under the curve was 0.83 with P value <0.001, the SWE test was good to regard as diagnostic test for risky pregnancy, with The cutoff value maximizing the accuracy of diagnosis was 1.27 m/s, sensitivity, specificity of this cutoff value were 82.5%, 67% respectively, Fig 2.



Fig 2: ROC curve for Placental SWE values showing cut of value 1.27 m/s with sensitivity of 82% and specificity



Fig 3: 30 Years' pregnant patient, gestational age about 30 weeks, placental thickness 3.8cm, ROI is placed in the center of colored area with velocity value measured as 1.07 m/s



Fig 4: 35 Years old patient with gestational DM, gestational age about 34weeks, placental thickness 3.7cm, ROI is placed in the center of colored area with velocity value 1.26 m/s

Discussion

Shear Wave Elastography (SWE), a non-invasive method assessing tissue stiffness, is advantageous in prenatal care due to its non-operator dependency and ability to provide additional functional information compared to B-mode and Doppler ultrasonography. While Doppler US has been utilized to predict preeclampsia by inspecting uterine artery notches and pulsatility index, SWE offers another dimension by measuring placental stiffness-a marker less studied but potentially indicative of pathology. This study is pioneering in using SWE to assess placental elasticity during the second and third trimesters of pregnancy. It found that women with gestational hypertension presented with the highest mean SWE values, corroborating Ohmaru et al.'s findings ^[6]. Similarly, women with gestational diabetes also showed higher mean SWE values, aligning with results from Yuskel et al. [14], suggesting that placental stiffness could be linked to specific pathologies like villous immaturity and chorangiosis observed in diabetic pregnancies. Although placental thickness didn't differ significantly between normal and high-risk pregnancies, a strong correlation with SWE was observed in high-risk cases, supporting findings by Altunkeser et al. [21]. This might be due to underlying conditions such as placental infarction or inflammation, more common in preeclampsia and diabetes. The study also reported a positive correlation between amniotic fluid index (AFI) and SWE in high-risk pregnancies, a finding that resonates with Khanal et al. [22] and Edward et al. ^[23]. However, no significant correlation between SWE values with maternal age or obstetric history was found, which is consistent with previous research ^[21]. For the identification of high-risk pregnancies, a cutoff value of 1.27 m/s was determined with sensitivity and specificity rates of 82.5% and 67%, respectively. Due to the low specificity, there's a risk of false positives; hence, additional testing, such as uterine artery Doppler flow velocimetry, could be employed for confirmation. Other studies, like Hefeda et al.^[24] and Fujita et al.^[25], found different cutoff values for predicting complications, which may be due to varied inclusion criteria and gestational age at examination.

Conclusion

SWE has shown to be a valuable diagnostic method for assessing placental health, especially in pregnancies at risk. It was observed that the mean SWE values were markedly higher in those with gestational hypertension compared to normal pregnancies. Additionally, a strong correlation was found between SWE, placental thickness, and amniotic fluid index in pregnancies deemed high-risk. For diagnosing such at-risk pregnancies, an SWE cutoff value of 1.27 m/s was determined to be most accurate, yielding a sensitivity of 82.5% and a specificity of 67%.

Conflict of Interest

Not available

Financial Support

Not available

References

1. Brosens I, Pijnenborg R, Vercruysse L, Romero R. The Great Obstetrical Syndromes are associated with disorders of deep placentation. American journal of obstetrics and gynecology. 2011;204(3):193-201.

- 2. Carter AM, Enders AC. Comparative aspects of trophoblast development and placentation. Reproductive Biology and Endocrinology. 2004;2(1):1-15.
- 3. Ami O, Maran J-C, Musset D, Dubray C, Mage G, Boyer L. Using Magnetic Resonance Imaging During Childbirth to Demonstrate Fetal Head Moldability and Brain Compression: Prospective Cohort Study. JMIR Formative Research. 2022;6(11):e27421.
- Sigrist RM, Liau J, El Kaffas A, Chammas MC, Willmann JK. Ultrasound elastography: review of techniques and clinical applications. Theranostics. 2017;7(5):1303.
- 5. Sugitani M, Fujita Y, Yumoto Y, Fukushima K, Takeuchi T, Shimokawa M, *et al.* A new method for measurement of placental elasticity: acoustic radiation force impulse imaging. Placenta. 2013;34(11):1009-13.
- 6. Ohmaru T, Fujita Y, Sugitani M, Shimokawa M, Fukushima K, Kato K. Placental elasticity evaluation using virtual touch tissue quantification during pregnancy. Placenta. 2015;36(8):915-20.
- Wu S, Nan R, Li Y, Cui X, Liang X, Zhao Y. Measurement of elasticity of normal placenta using the Virtual Touch quantification technique. Ultrasonography. 2016;35(3):253.
- 8. Wang M, Byram B, Palmeri M, Rouze N, Nightingale K. On the precision of time-of-flight shear wave speed estimation in homogeneous soft solids: initial results using a matrix array transducer. IEEE transactions on ultrasonics, ferroelectrics, and frequency control. 2013;60(4):758-70.
- 9. Shiina T. WFUMB guidelines and recommendations for clinical use of ultrasound elastography: Part 1: basic principles and terminology. Ultrasound in Medicine and Biology. 2017;43:S191-S2.
- 10. Cosgrove DO, Berg WA, Doré CJ, Skyba DM, Henry J-P, Gay J, *et al.* Shear wave elastography for breast masses is highly reproducible. European radiology. 2012;22(5):1023-32.
- 11. Dietrich CF, Bamber J, Berzigotti A, Bota S, Cantisani V, Castera L, *et al.* EFSUMB guidelines and recommendations on the clinical use of liver ultrasound elastography, update 2017 (Long version). Ultraschall in der Medizin-European Journal of Ultrasound. 2017;38(04):e16-e47.
- 12. Regnault T, Galan H, Parker T, Anthony R. Placental development in normal and compromised pregnancies-a review. Placenta. 2002;23:S119-S29.
- 13. Rouze NC, Wang MH, Palmeri ML, Nightingale KR. Parameters affecting the resolution and accuracy of 2-D quantitative shear wave images. IEEE transactions on ultrasonics, ferroelectrics, and frequency control. 2012;59(8):1729-40.
- 14. Yuksel MA, Kilic F, Kayadibi Y, Alici Davutoglu E, Imamoglu M, Bakan S, *et al.* Shear wave elastography of the placenta in patients with gestational diabetes mellitus. Journal of Obstetrics and Gynaecology. 2016;36(5):585-8.
- 15. Shiina T. JSUM ultrasound elastography practice guidelines: basics and terminology. Journal of Medical Ultrasonics. 2013;40(4):309-23.
- 16. Thiele M, Madsen BS, Procopet B, Hansen JF, Møller LMS, Detlefsen S, *et al.* Reliability criteria for liver

stiffness measurements with real-time 2D shear wave elastography in different clinical scenarios of chronic liver disease. Ultraschall in der Medizin-European Journal of Ultrasound. 2017;38(06):648-54.

- 17. Issaoui M, Debost-Legrand A, Skerl K, Chauveau B, Magnin B, Delabaere A, *et al.* Shear wave elastography safety in fetus: a quantitative health risk assessment. Diagnostic and interventional imaging. 2018;99(9):519-24.
- Li C, Zhang C, Li J, Cao X, Song D. An experimental study of the potential biological effects associated with 2-D shear wave elastography on the neonatal brain. Ultrasound in Medicine & Biology. 2016;42(7):1551-9.
- 19. Shiina T, Nightingale KR, Palmeri ML, Hall TJ, Bamber JC, Barr RG, *et al.* WFUMB guidelines and recommendations for clinical use of ultrasound elastography: Part 1: basic principles and terminology. Ultrasound in medicine & biology. 2015;41(5):1126-47.
- Ter Haar G. The new British Medical Ultrasound Society Guidelines for the safe use of diagnostic ultrasound equipment. SAGE Publications Sage UK: London, England; c2010. p. 50-1.
- 21. Altunkeser A, Alkan E, Günenç O, Tolu I, Körez MK. Evaluation of a healthy pregnant placenta with shear wave elastography. Iranian Journal of Radiology. 2019, 16(1).
- 22. Khanal UP, Chaudhary R, Ghanshyam G. Placental elastography in intrauterine growth restriction: a case–control study. Journal of clinical research in radiology. 2019;2(2):1-7.
- 23. Edwards C, Cavanagh E, Kumar S, Clifton VL, Borg DJ, Priddle J, *et al.* Changes in placental elastography in the third trimester-analysis using a linear mixed effect model. Placenta. 2021;114:83-9.
- 24. Hefeda MM, Zakaria A. Shear wave velocity by quantitative acoustic radiation force impulse in the placenta of normal and high-risk pregnancy. Egyptian journal of radiology and nuclear medicine. 2020;51:1-12.
- 25. Fujita Y, Nakanishi TO, Sugitani M, Kato K. Placental elasticity as a new non-invasive predictive marker of pre-eclampsia. Ultrasound in medicine & biology. 2019;45(1):93-7.

How to Cite This Article

Najee FK, AL-Tameemi HN. Evaluation of placental stiffness in normal versus high risk pregnancy using shear wave elastography. International Journal of Radiology and Diagnostic Imaging 2023; 6(4): 08-13.

Creative Commons (CC) License

This is an open access journal, and articles are distributed under the terms of the Creative Commons Attribution-NonCommercial-ShareAlike 4.0 International (CC BY-NC-SA 4.0) License, which allows others to remix, tweak, and build upon the work non-commercially, as long as appropriate credit is given and the new creations are licensed under the identical terms.